



DOES *Beta vulgaris* L. HAVE *LCYB* GENE?

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ABSTRACT

Beta carotene is a pigment that generally shared by all plants. This pigment synthesis is catalyzed by lycopene beta cyclase enzyme. This enzyme is encoded by the *LCYB* gene. This research aimed to obtain the sequence of *LCYB* gene isolated from beet (*Beta vulgaris* L.) leaves. Several experiments have been done to isolate the *LCYB* gene. In the first experiment, the gene primers are designed using Pick Primer software based on conserved region of *Taraxacum officinale* (accession no. AB247456.1) and *Lilium lancifolium* *LCYB* (accession no. GU471230.1). The forward primer is (5'-TGTCGTGGTGGATCTTGTGG-3') and the reverse is (5'-ACACCTGTTGAGCGACAGAC-3'). At the first experiment, the gene target's sequence was not obtained as the phylogenetic relationship between *Beta vulgaris*, *Taraxacum officinale*, and *Lilium lancifolium* were too far. In the second experiment, the primers are designed using *LCYB* gene in *Arabidopsis thaliana* (accession no. NM_111858.2), with considering *Arabidopsis thaliana* as a plant model. The primers are (5'-TGGGACAGCAGGAATGGTTC-3') as the forward primer and (5'-GAGAAGAGCGACAACCCGAA-3') the reverse. By using BLAST analysis in the second experiment, it suggests that the fragment amplified by forward primer has similarity with *trnL* gene. This study seems to amplify the *trnL* gene instead of *LCYB* gene. This fact leads to bigger question whether or not the *Beta vulgaris* L has the *LCYB* gene.

Key words : Beta carotene, Lycopene Beta Cyclase, *Beta vulgaris* L, *LCYB* gene

INTRODUCTION

Carotenoids is a precursor in vitamin A biosynthesis (Giuliano *et al.*, 2008). This pigment is a C40 isoprene derived from the C20 precursor geranylgeranylpyrophosphate (GGPP) (Yamamoto *et al.*, 2009). Deficiency of vitamin A resulted on night blindness (Ye *et al.*, 2000), which in turn stimulates the research on plant breeding in order to produce various plants with high carotenoid content (provitamin A) (Giuliano *et al.*, 2008). Beet is one of plants which contains high carotenoids (Berman *et al.*, 2004). Genetic engineering affects the expression of genes encoding carotenoid pigments that have an impact on levels of carotene (Bai *et al.*, 2011; Giuliano *et al.*, 2008). Beta-carotene, one of carotenoids pigments (Brown, 2010) is converted by *Lycopene Beta Cyclase* enzyme. The enzyme is encoded by the *Lycopene Beta Cyclase (LCYB)* gene (Cunningham *et al.*, 1998). A report about *LCYB* gene sequence in *Beta vulgaris* L. has not yet been available. This research aimed to isolate *LCYB* from beet, yet resulted on a question does *Beta vulgaris* L. has *LCYB* gene?

MATERIALS AND METHODS

In this study, DNA were isolated using Nucleospin®II DNA Isolation Kit, (Macherey-Nagel, Germany). In the first experiment the primer *LCYB* gene were designed using PickPrimer software based on conserved region of *Taraxacum officinale* (accession no. AB247456.1) for forward primer (5'-TGTCGTGGTGGATCTTGTGG-3') and *Lilium lancifolium* (accession no. GU471230.1) for reverse primer (5'-ACACCTGTTGAGCGACAGAC-3'). Since first

study resulted an unspecific sequence we therefore continued the study with second experiment using primers for *LCYB* gene which was designed based on conserved region of *Arabidopsis thaliana LCYB* (accession no.NM_111858.2).

The forward primer (5'-TGGGACAGCAGGAATGGTTC-3') and reverse primer (5'-GAGAAGAGCGACAACCCGAA-3'). PCR results were examined using electrophoresis gel agarose in 1.5% and 1Kbp DNA Marker. The data was analysed using BLAST, Clustal X and BioEdit software.

RESULTS AND DISCUSSION

Total DNA *B.vulgaris* L.obtained was of 245.6 ng/μL. The target gene was amplified by PCR through several stages of annealing temperature optimizations. The first experiment produced a thin band of 200 bp fragment which showed a nonspecific sequences for both forward and reverse primer (Figure 1). Analysis using DNA Base did not showed a consensus sequence from forward and reverse fragment (Figure 2). Using BLAST analysis the forward fragment has query coverage of 6% with maximum identity 100% to *Taraxacum officinale LCYB* while the reverse fragment has a query coverage of 12% and maximum identity 100% to *Lilium lancifolium LCYB*. We suggested that the failure to obtain the target gene was generated primer by miss-design since later we find that *B. vulgaris* L. too far from *Taraxacum officinale* and *Lilium lancifolium* phylogenetically (Graur *et al.*, 2000).

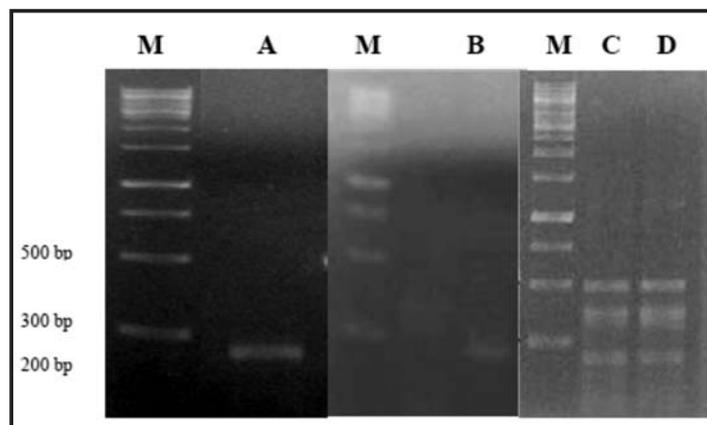


Figure1. Results of PCR at annealing temperature A. 61°C (200 bp); B. 57°C (200 bp); C. 52 °C (200 bp, 400 bp, and 500 bp); D.55°C (200 bp, 400 bp, and 500 bp);M=Marker 1 kbp DNA

Based on that result we performed the second experiment which resulted a 200bp fragment, 400bp and 500bp(GeneRuler 1kbp on agarose gel 1.5% (Figure 1).Analysis using DNA Baser did not obtained a consensus between forward and reverse (Figure 3). BLAST analysis showed DNA fragments obtained are not specific. The forward sequence (AF) showed similarity to *trnL* with a maximum identity 98% and query coverage 34% of *B. vulgaris* L. This failure may be caused by primers design failure as a result of the lack conserved region of *LCYB* gene. Another possibility was based on the fact that carotenoid biosynthesis in different plants is catalyzed by not only a single type of enzyme, yet a family proteins and different genes.

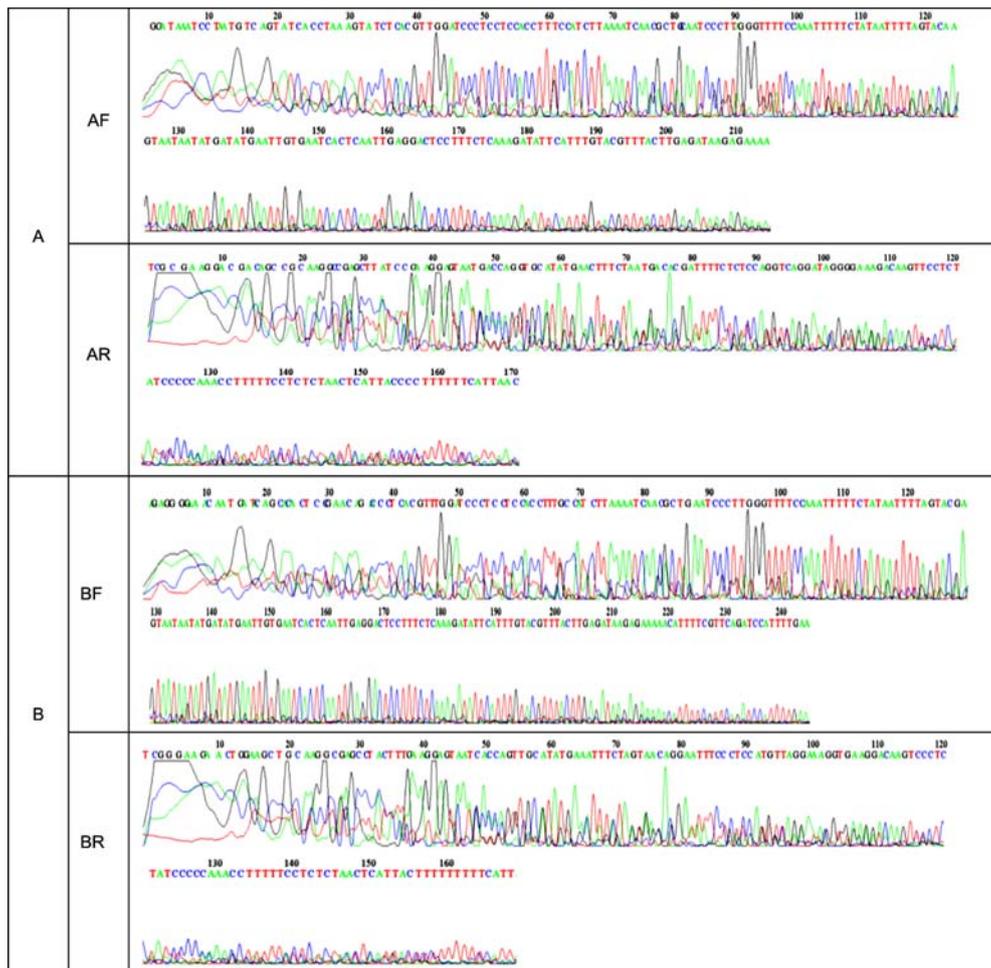


Figure3. The results of the second experiment sequencing; A. Fragment 500 bp (AF: forward, AR: reverse); B. Fragment 400 bp (BF: forward, BR: reverse).

REFERENCES

- Alquézar, B., L. Zacarias, and M.J. Rodrigo. 2009. Molecular and Functional Characterization of A Novel Chromoplast-Specific Lycopene β -Cyclase from Citrus and its Relation to Lycopene Accumulation. *Journal Experimental Botany* 60: 1783-1797.
- Altschul, S. F., W. Gish, W. Miller, E.W. Myers, and D.J. Lipman. 1990. Basic Local Alignment Search Tool. *J. Mol. Biol.* 215: 403-410.
- Bai, C., R.M. Twyman, G. Farre, G. Sanahuja, P. Christou, T. Capell, and C. Zhu. 2011. A golden era pro vitamin A enhancement in diverse crops. *In Vitro Cell Dev. Biol. Plant* 47: 205-221.
- Berman, A. F., M.J. Balick, F. Kronenberg, A.L. Ososki, B. Connor, M. Reiff, M. Roble, P. Lohr, B.J. Brosi, and R. Lee. 2004. Treatment of fibroids: the use of beets (*Beta vulgaris*) and molasses (*Saccharum officinarum*) as an herbal therapy by Dominican healers in New York City. *Journal of Ethnopharmacology* 92: 337–339.
- Brown, J. L. 2010. *Supplement Facts Vitamin A*. Pennsylvania: Pennsylvania State University.
- Graur, D., and W. Li. 2000. *Fundamentals of Molecular Evolution (Second Edition)*. Sunderland: Sinauer Associates, Inc.

- Cunningham Jr, F. X. , and E. Gant. 1998. Genes and Enzymes of Carotenoid Biosynthesis in Plants. *Plant Physiology and Plant Molecular Biology* 49: 557-583.
- Giuliano, G., R. Tavazza, G. Diretto, P. Beyer, and M.A. Taylor. 2008. Metabolic engineering of carotenoid biosynthesis in plants. *Cell Press* 3: 0167-7799. Intron Biotechnology.
2012. *I-Taq™ DNA Polymerase*. Intron Biotechnology, Inc.
- McPherson, M. J., and S.G. Moller. 2006. *PCR (Second Edition)*. New York : Taylor & Francis Group.
- Ruiz-Sola, M. A., and M. Rodríguez-Concepcion. 2012. Carotenoid Biosynthesis in Arabidopsis: A Colorful Pathway. American Society of Plant Biologists.
- Yamamizo, C., S. Kishimoto, and A. Ohmiya. 2009. Carotenoid composition and carotenogenic gene expression during *Ipomoea* petal development. *Journal of Experimental Botany*. 10: 1-11.
- Ye, X., S.A. Babili, A. Kloti, J. Zhang, P. Lucca, P. Beyer, and I. Potrykus. 2000. Engineering the Provitamin A (β -Carotene) Biosynthetic Pathway into (Carotenoid Free) Rice Endosperm. *Science* 287: 303-305.