

Research article

Antioxidant Activities of Green Tea (Camellia Sinensis L.) Leaves From Ciwidey, West Java

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Abstract.

It has been reported that polyphenols-rich diets provide some protection against the development of cancers, cardiovascular diseases, diabetes, osteoporosis and neurodegenerative diseases. Exogenous antioxidants are abudantly found in plants. Tea (*Camellia sinensis* L.; family *Theaceae*), which is the most popular beverage in Southeast Asia, is known for its medicinal properties. In this study, we evaluated antioxidant activity as well as phenol and flavonoid content in green tea leaves from PPTK Gambung Ciwidey in West Java. Antioxidant activity was observed through measuring 2,2- diphenyl-1 picrylhydrazyl (DPPH) scavenging activity. The total phenolic content was determined by the Folin Ciocalteu method. The green tea leaf extract showed the highest DPPH-scavenging activity at a concentration of 25 ug/ml (DPPH = 94.55%;IC₅₀ = 0.54 µg/mL). Furthermore, the total phenolic and flavonoid contents of the green tea leaf extract were 36.64 and 8.43 mg QE/mg extract, respectively. These findings showed that the green tea leaf extract exhibited antioxidant activity.

Keywords: Camellia sinensis L, flavonoid, antioxidant

1. Introduction

Free radicals are biochemical responses in the body which further promotes cancer, ischemic heart disease, inflammation, diabetes, aging, atherosclerosis, immunosuppression, and neurodegenerative disorders. [1-3] Antioxidant system in human body is usually responsible to scavenge the radicals. However, this might be interferred by excessive *reactive oxygen species* (ROS) and *reactive nitrogen species* (RNS) due to the exposure of cigarette smoking, alcohol, radiation, or environmental toxins. [2, 4, 5] Exogenous antioxidants are considered to restore such event by inhibiting oxidative chain reaction [6].

The exogenous antioxidants are mostly derived from food and medicinal plants [7-10]. These natural antioxidants are mainly polyphenols (phenolic acids, flavonoids, anthocyanins, lignans and stilbenes), carotenoids (xanthophylls and carotenes) and vitamins (vitamin E and C) [6]. Polyphenols and carotenoids exhibit many biological

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activities, such as anti-inflammatory, antibacterial, antiviral, anti-aging, and anticancer properties. [11-13]

Antioxidant activities have been widely studied on numerous plants. Tea (*Camellia sinensis* L.; family Theaceae), a well-known beverage in Southeast Asia, has been acknowledged for its medicinal properties [14-16]. White, green, oolong, black, and pu'erh teas are the major tea types sourced from leaves and buds of the tea plant [17]. These tea types also differ based on the variety of *C. sinensis* used in their production. Some studies support that among all tea types, green teas contain the highest amount of catechins, a group of polyphenolic flavan-3-ol monomers and their gallate derivatives [18]. The major catechins include (–)-epicatechin (EC), (–)- epigallocatechin (EGC), (–)-epicatechin-3-gallate (ECG), and (–)-epigallocatechin-3-gallate (EGCG). These compounds are responsible for its various biological functions, including antioxidant activities, anti-inflammatory activities, anti-melanogenic effects, and hepatoprotection [19-23]. Similar polyphenolic content among green tea varieties can provide more options of healthy beverage for people. Thus, we aimed to evaluate phenol and flavonoid content as well as antioxidant activity of green tea leaves from PPTK Gambung Ciwidey West Java.

2. Materials and Method

2.1. Green tea extraction

Oolong tea (C. sinensis) was obtained from a tea plantation in East Java. Oolong tea was extracted with 96% methanol (1:4) using a maceration technique. The filtrate was filtered with Na_2SO_4 and collected every 24 h until the filtrate became colorless. The filtrate was evaporated at $40^{\circ}C$ in an evaporator until a dried pellet was obtained. The ethanol extracted pellet was stored at $4^{\circ}C$ [24].

2.2. Fractination (Thin-layer Chromatography)

Thin-layer chromatography is an early identification of flavonoid. Extract was dissolved in three solvents: water, ethyl acetate, and n-hexane. Solution was filtered, and then evaporated. End product was spotted onto KLT plate, and eluted with mixture of water:ethyl acetate:n-hexane. Dots were observed and Rf was measured [24].



2.3. Total phenolic content

Total phenolic content was determined by Folin Ciocalteu method. Folin Ciocalteu's reagent (10%, 75 µl) was added to the samples (15 µl). Sodium carbonate solution (7,5 %, 60 µl) was then added to the samples, whilts DMSO (135 µl) was added into blank. Plates were incubated for 10 min in the dark at 50°C. The absorbance was measured at 760 nm wavelength. The total phenolic content was expressed as gallic acid equivalents (GAE) mg/g of tea leaves.

$$Total \ phenolic \ content = \frac{Sample \ absorbance \ -}{a}$$

b

v = ax + b

2.4. Measurement of flavonoid

Green tea leaves extract and guercetin (75 µl) was added into samples and blank well. Alumunium chloride (2%, 75 µl) was added into samples, whilst DSMO (75 µl) was added into blank well. Absorbance was recorded at 415 nm wavelength. Flavonoid total was measured according to linier regression of quercetin.

$$y = ax + b$$

Total flavonoid = $\frac{Sample \ absorbance - b}{a}$

2.5. DPPH (2,2- diphenyl-1 picrylhydrazyl) scavenging activity

Briefly, 50 µl extracts and eugenol (Sigma-Aldrich) were added to a microplate followed by 200 µl DPPH (Sigma-Aldrich) solution (0.077 mmol/l in methanol). The mixtures was shaken vigorously and kept in the dark for 30 min at room temperature; DPPH scavenging activity was determined with a microplate reader at 517 nm. Negative control was 50 μ L DMSO and 200 μ L DPPH, and blank was 50 μ L sample and 200 μ L DMSO 10% [25]. The radical scavenging activity of each sample was measured according to following formula:

$$DPPH - scavenging \ activity = 1 - \frac{Samples \ absorbance}{Negative \ control \ absorbance} \times 100$$



3. Results

Phenolic content was measured with Folin-Ciocalteu colorymetry method in which polyphenol of plants reacts to redox specific reagent to form blue complex [26]. As shown in Table 3, phenolic total of green tea leaves extract was $36,64 \pm 0,39$ mg GAE/ mg extract.

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|-------------------|--------------|----------------|-----------------|
| TABLE 1: Analysis | or prienolic | content of tea | leaves extract. |

| Sample | Phenolic content (GAE/mg extract) | (mg | Flavonoid content (1 mg QE/1 mg sample) |
|-----------------------------|---------------------------------------|-----|--|
| Green tea leaves extract | 36,64 <u>+</u> 0,39 | | 8,43±0,39 |

Flavonoid content was measured with complementary colorimetry of yellow complex formed by reaction between $AICI_3$ and flavonols [27]. Total flavonoid of green tea leaves extract obtained in this study (Table 3) was 8,43 \pm 0,39 mg QE/mg extract.

| Green tea leaves extract (ug/ml) | DPPH-scavenging (%) | activity | IC ₅₀ (μg/mL) |
|-------------------------------------|-------------------------------|----------|--------------------------|
| 25,0 | 94,55 \pm 0,78 ^e | | 0,54 ± 0,08 |
| 12,5 | $75,96 \pm 0,40^{d}$ | | |
| 6,25 | 64,72 ± 0,17 ^c | | |
| 3,13 | 54,08 ± 0,74 ^b | | |
| 1,56 | 48,04 ± 0,16 ^a | | |

TABLE 2: DPPH-scavenging activity of gree tea leaves extract.

Data are presented in Mean \pm Standard Deviation. Superscript letters ^{*a*-*e*} in each column indicates signicant difference (p < 0,05) (Tukey HSD post hoc test).

DPPH-scavenging activity and IC₅₀ value of green tea leaves extract are presented in Table 1. Green tea leaves extract showed highest DPPH-scavenging activity (94,55 \pm 0,78%). The IC₅₀ of the extract was 0,54 \pm 0,08 µg/mL.

4. Discussion

In the present study, green tea leave extract exhibited antioxidant properties as shown in high DPPH-scavenging activity. The test with the DPPH radical aims at labeling the ability of its neutralization via the antioxidants included in the solution [28]. The result of presents study showed highest DPPH-scavenging activity of green tea leaves extract, was at concentration of 25 ug/ml (DPPH = 94,55%⁻IC₅₀ = 0,54 µg/mL). This was higher than previous study done by Nazliniwaty, et al (2021) in which C. sinensis extract exhibits lower antioxidant activity (IC50 = 11.83 \pm 0.005 µg/ml) [29]. KnE Medicine



Antioxidant properties are associated with presence of polyphenols, flavanoids and epigallocatechin gallate in tea leaves [30]. Polyphenols are secondary metabolites of plants generally involved in defense against ultraviolet radiation, aggression by pathogens [31], and stress [32]. In the present study, total phenolic content and flavonoid of green tea leaves extract were 36,64 and 8,43 mg QE/mg extract respectively Previous study on green tea from Argentina shows total polyphenol was ranged from 21.02 +/- 1.54 to 14.32 +/- 0.45% of gallic acid equivalents (GAE) [33]. In addition, Almajano et al. (2008) revealed that total phenolic content in green tea was higher than black tea [34].

Different polyphenol content among tea varieties might be infuenced by several factors such as degree of ripeness, environmental factors, processing and storage [35]. Maturation of tea leaves influence polyphenols content and antioxidant activities due to the morphological changes and chemical compounds transportation within the plant [36]. Levels of EGCC and ECG was found higher in young tea leaves than mature leaves [37]. In contrast, study carried by Lin *et al.* (2003) shows that EGCG, EGC, EC and catechin are richer in old leaves than in young leaves [18]. Different solvents also plays role in antioxidant activities. Solvent that has higher polarity (aqueous-methanol), is more efficient to scavenge free radicals than less polar solvent (methanol and hot water) [38].Ethanol is frequently considered as an efficient solvent to extract polyphenols [39].

Although there are many studies regarding the promising medicinal properties of green tea in Asia, we believe that this study is new addition from Indonesian plants, especially from West Java region, which can be further compared to other tea variants. We plan to optimize and to explore more activities from green tea in the near future.

5. Conclusion

Green tea leave extract exhibited antioxidant activity which is associated with its phenolic content. Further in vitro and in vivo studies to observe the efficacy of green tea leave extract, are encouraged.

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References

- [1] Peng C, Wang X, Chen J et al. Biology of ageing and role of dietary antioxidants. BioMed Research International. 2014:1(1):1-13.
- [2] Li S, Tan HY, Wang N. The role of oxidative stress and antioxidants in liver diseases. International Journal of Molecular Sciences. 2015;16(11):26087-26124.
- [3] Chanda S, Dave R, Kaneria M. In vitro antioxidant property of some Indian medicinal plants. Reserach Journal Medicine Plants. 2011;5(2):169-179.
- [4] Wang F, Li Y, Zhang YJ, Zhou Y, Li S, Li HB. Natural products for the prevention and treatment of hangover and alcohol use disorder. Molecules. 2016;21(1):64-80.
- [5] Zhou Y, Zheng J, Li S, Zhou T, Zhang P, Li HB. Alcoholic beverage consumption and chronic diseases. International Journal of Environmental Research and Public Health. 2016;13(6):522-539.
- [6] Baiano A, Del Nobile MA. Antioxidant compounds from vegetable matrices: Biosynthesis, occurrence, and extraction systems. Critical Reviews in Food Science and Nutrition. 2016;56(12):2053-2068.
- [7] Deng GF, Lin X, Xu XR, Gao LL, Xie JF, Li HB. Antioxidant capacities and total phenolic contents of 56 vegetables. Journal of Functional Foods. 2013;5(1):260-266.
- [8] Li AN, Li S, Li HB, Xu DP, Xu XR, Chen F. Total phenolic contents and antioxidant capacities of 51 edible and wild flowers. Journal of Functional Foods. 2014;6:319-330.
- [9] Li S, Li SK, Gan RY, Song FL, Kuang L, Li HB. Antioxidant capacities and total phenolic contents of infusions from 223 medicinal plants. Industrial Crops and Products. 2013;51:289-298.
- [10] Li Y, Zhang JJ, Xu DP, Zhou T, Zhou Y, Li S, Li HB. Bioactivities and health benefits of wild fruits. International Journal of Molecular Sciences. 2016;17(8):1258-1285.
- [11] Zhang YJ, Gan RY, Li S et al. Antioxidant phytochemicals for the prevention and treatment of chronic diseases. Molecules. 2015;20(12):21138-21156.
- [12] Prasad KN. Simultaneous activation of Nrf2 and elevation of antioxidant compounds for reducing oxidative stress and chronic inflammation in human Alzheimer's disease. Mechanisms of Ageing and Development. 2016;153:41-47.
- [13] Zhou Y, Zheng J, Li Y et al. Natural polyphenols for prevention and treatment of cancer. Nutrients. 2016;8(8):515-550.
- [14] Henning SM, Niu Y, Liu Y et al. Bioavailability and antioxidant effect of epigallocatechin gallate administered in purified form versus as green tea extract in healthy individuals. The Journal of Nutritional Biochemistry. 2005;16(10):610-616.



- [15] Camargo LEA, Pedroso LS, Vendrame SC, Mainardes RM, Khalil NM. Antioxidant and antifungal activities of *Camellia sinensis* (L.) kuntze leaves obtained by different forms of production. Brazilian Journal of Biology. 2016;76:428-434.
- [16] Gramza-Michałowska A, Kobus-Cisowska J, Kmiecik D et al. Antioxidative potential, nutritional value and sensory profiles of confectionery fortified with green and yellow tea leaves (*Camellia sinensis*). Food Chemistry. 2016;211:448-454.
- [17] Pettigrew J. The tea companion: A connoisseur's guide. 1st ed. Philadelphia: Running Press Book Publishers; 2004.
- [18] Lin Y, Tsai Y, Tsay J, Lin J. Factors affecting the levels of tea polyphenols and caffeine in tea leaves. Journal Agric Food Chemistry. 2003;51:1864–73
- [19] Tipoe GL, Leung TM, Hung MW, Fung ML. Green tea polyphenols as an anti-oxidant and anti-inflammatory agent for cardiovascular protection. Cardiovascular & Haematological Disorders-Drug Targets (Formerly Current Drug Targets-Cardiovascular & Hematological Disorders). 2007;7(2):135-144.
- [20] Kim YC, Choi SY, Park EY. Anti-melanogenic effects of black, green, and white tea extracts on immortalized melanocytes. Journal of Veterinary Science. 2015;16(2):135-143.
- [21] El-Beshbishy HA. Hepatoprotective effect of green tea (*Camellia sinensis*) extract against tamoxifen-induced liver injury in rats. BMB Reports. 2005;38(5):563-570.
- [22] Issabeagloo E, Taghizadieh M. Hepatomodulatory action of *Camellia sinensis* aqueous extract against isoniazid-rifampicin combination induced oxidative stress in rat. Advances in Bioresearch. 2012;3(3):401-415.
- [23] Lodhi P, Tandan N, Singh N, Kumar D, Kumar M. Camellia sinensis (L.) kuntze extract ameliorates chronic ethanol-induced hepatotoxicity in albino rats. Evidence-Based Complementary and Alternative Medicine. 2014:1(2):1-5.
- [24] Hostettmann K. Handbook of chemical and biological plant analytical methods. John Wiley & Sons; Unites States. 2014.
- [25] Widowati W, Herlina T, Ratnawati H, Constantia G, Deva IDGS, Maesaroh M. Antioxidant potential of black, green and oolong tea methanol extracts. Biology, Medicine, & Natural Product Chemistry. 2015;4(2):35-39.
- [26] Schofield P, Mbugua DM, Pell AN. Analysis of condensed tannins: A review. Animal Feed Science and Technology. 2001;91(1-2):21-40.
- [27] Chang CC, Yang MH, Wen HM, Chern JC. Estimation of total flavonoid content in propolis by two complementary colorimetric methods. Journal of Food and Drug Analysis. 2002;10(3):178-182.



- [28] Koss-Mikołajczyk I, Baranowska M, Namie´snik J, Bartoszek A. Determination of antioxidant activity of phytochemicals in cellular models by fluorescence/luminescence methods. Postepy Higieny i Medycyny Doswiadczalnej. 2017;71:602–617.
- [29] Nazliniwaty T, Laila L. Antioxidant activity test of green tea (*Camellia sinensis* L. kuntze) ethanolic extract using DPPH Method. Science and Technology publication. Indonesia. 2020.
- [30] Fernando CD, Soysa P. Extraction kinetics of phytochemicals and antioxidant activity during black tea (*Camellia sinensis* L.) brewing. Nutrition Journal. 2015;14(1):1-7.
- [31] Beckman CH. Phenolic-storing cells: Keys to programmed cell death and periderm formation in wilt disease resistance and in general defence responses in plants? Physiological and Molecular Plant Pathology. 2000;57(3):101-110.
- [32] Parr AJ, Bolwell GP. Phenols in the plant and in man. The potential for possible nutritional enhancement of the diet by modifying the phenols content or profile. Journal of the Science of Food and Agriculture. 2000;80(7):985-1012.
- [33] Anesini C, Ferraro GE, Filip R. Total polyphenol content and antioxidant capacity of commercially available tea (*Camellia sinensis*) in Argentina. Journal of Agricultural and Food Chemistry. 2008;56(19):9225-9229.
- [34] Almajano MP, Carbo R, Jiménez JAL, Gordon MH. Antioxidant and antimicrobial activities of tea infusions. Food Chemistry. 2008;108(1):55-63.
- [35] Manach C, Scalbert A, Morand C, Rémésy C, Jiménez L. Polyphenols: Food sources and bioavailability. The American Journal of Clinical Nutrition. 2004;79(5):727-747.
- [36] Farhoosh R, Golmovahhed GA, Khodaparast MH. Antioxidant activity of various extracts of old tea leaves and black tea wastes (*Camellia sinensis* L.). Food Chemistry. 2007;100(1):231-236.
- [37] Chen CN, Liang CM, Lai JR, Tsai YJ, Tsay JS, Lin JK. Capillary electrophoretic determination of theanine, caffeine, and catechins in fresh tea leaves and oolong tea and their effects on rat neurosphere adhesion and migration. Journal of Agricultural and Food Chemistry. 2003;51(25):7495-7503.
- [38] Turkmen N, Sari F, Velioglu YS. Effects of extraction solvents on concentration and antioxidant activity of black and black mate tea polyphenols determined by ferrous tartrate and Folin–Ciocalteu methods. Food Chemistry. 2006;99(4):835-841.
- [39] Koffi E, Sea T, Dodehe Y, Soro S. Effect of solvent type on extraction of polyphenols from twenty-three Ivorian plants. Journal of Animal and Plant Sciences (JAPS). 2010;5(3):550-558.