

Conference Paper

Pun1 Gene Isolation from *Capsicum frutescens* L. cv Cakra Hijau

Elhah Nailul Khasna, Shelly Zairina, Ria Reinnata Juliandari, Eko Sri Sulasmi, and Dwi Listyorini

Departement of Biology, Faculty of Mathematics and Natural Science, State University of Malang, Jl. Semarang 5, Malang, 651335, Indonesia

Abstract

Capsicum frutescens L. is one of chili peppers with high pungency. *Capsicum frutescens* has several cultivars, one of those is *C. frutescens* cv. Cakra Hijau. This cultivars is known resistance to pests and diseases as well. Pungency is due to the accumulation of capsaicinoids. *Pun1* is an important gene responsible for pungency. The full-length genomic sequence of *Pun1* is 1897 bp, containing two exons of 738 bp and 590 bp and one intron of 348 bp in between. This study was aimed to isolate *Pun1* gene that free from intron. mRNA was isolated with TriReagent furthermore RT-PCR method used Qiagent One-Step RT-PCR and two pairs of primer : F1/R1 (F15'-ATG-GCT-TTT-GCA-TTA-CCA-TCA-3'/R15'-CTT-AGC-TCG-AAG-TGC-ATC-TA-3') and F2/R2 (F25'-GAA-GGT-GGC-AGA-AGA-ATC-AG-3'/R25'-TTA-GGC-AAT-GAA-CTC-AAG-GA-3'). The result of this study are isolated 738 bp exon-1 and 590 bp exon 2 of *Pun1* gene.

Corresponding Author:

Elhah Nailul Khasna

elhahnailulhasna@gmail.com

Received: 11 February 2017

Accepted: 08 March 2017

Published: 26 March 2017

Publishing services provided
by Knowledge E

© Elhah Nailul Khasna et al. This article is distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use and redistribution provided that the original author and source are credited.

Selection and Peer-review under the responsibility of the ICBS Conference Committee.

Keywords: Capsaicin; *Capsicum frutescens* L. cv. Cakra Hijau; exon; *Pun1* gene.

1. Introduction

Capsicum frutescens L. is widely cultivated in tropical and subtropical regions of the world including Indonesia [1, 2]. Cakra Hijau is one of local cultivar from *C. frutescens* that have economic value, it is resistant to pest and disease as well and has high level of pungency [3]. The pungency level is due to the accumulation of capsaicinoids [4]. Capsaicinoid are synthesized and accumulated in the placental tissue [5]. Capsaicinoid was first detected at 20th days after anthesis then gradually increased and reached maximal level around 40 d after anthesis, on the 50 d, the capsaicinoid content decreased significantly [6]. Capsaicin synthase (CS) is the last enzyme in the biosynthesis of capsaicin and have important role in condensation of vanillylamine from phenylpropanoid with 8-methyl-6-nonenic acid from valine pathway [7]. The CS expression starts at 10 dpa, reaches a maximum at 20 dpa, then suddenly decreases and almost stops at 40 dpa [8]. *Pun1* gene encodes CS enzyme, this gene probably functions as an acyltransferase to complete the capsaicinoid synthesis. Non-pungent were observed to have a 2,5 kb deletion spanning the putative promoter and exon-1

OPEN ACCESS

of *Pun1* gene and a 4 bp deletion in part of exon-1 which disturb translation and so deletion of part of exon-2 including the stop codon [9–11].

DNA isolation of *Pun1* gene from *C. frutescens* cv. Cakra Hijau had been reported [12, 13]. This study aimed to isolate coding region from RNA of *Pun1* gene *C. frutescens* cv. Cakra Hijau to complete the existing data.

2. Materials and Methods

2.1. Plant material

Seeds of *Capsicum frutescens* cv. Cakra Hijau were obtained from Balai Pengkajian Teknologi Pertanian (BPTP) [Assesment Institute for Agricultural Technology—AIAT], Karang Ploso, Malang, Indonesia.

2.2. RNA extraction

Total RNA was extracted from placental tissue approximately 100 mg for each reaction using TriReagent (Invitrogen) according to the manufactures instruction. Quantity of RNA was measured using NanoDrop DR2000.

2.3. Reverse Transcriptase-PCR

Complementary DNA (cDNA) was synthesized using Qiagen OneStep RT-PCR (Qiagen) as per the manufacturer's instruction. In all, 500 ng of total RNA was used for each reaction. PCR primer were design based on *C. frutescens* cv. Shuanla sequence from Gene Bank (accession number HM854860). The primer used were F1/R1 (F15'-ATG-GCT-TTT-GCA-TTA-CCA-TCA-3'/R15'-CTT-AGC-TCG-AAG-TGC-ATC-TA-3') and F2/R2 (F25'-GAA-GGT-GGC-AGA-AGA-ATC-AG-3'/R25'-TTA-GGC-AAT-GAA-CTC-AAG-GA-3'). RT-PCR cycles a is follows: reverse transcription 50°C for 30 min, initial PCR activation step 95°C for 15 min, denaturation 94°C for 1 min, annealing 54°C for 1 min, extension 70°C for 1 min, and final extension 70°C for 10 min. RT-PCR product were checked using 1.5% gel electrophoresis and sequenced by ABIPRISM 3730x1 Genetic analyzer machine at First Base Laboratories, Malaysia.

2.4. Analysis

The sequence was analyzed using DNA Baser, BLASTN, Clustal X and SIXFRAME (as it is an online programe so please add the website address).

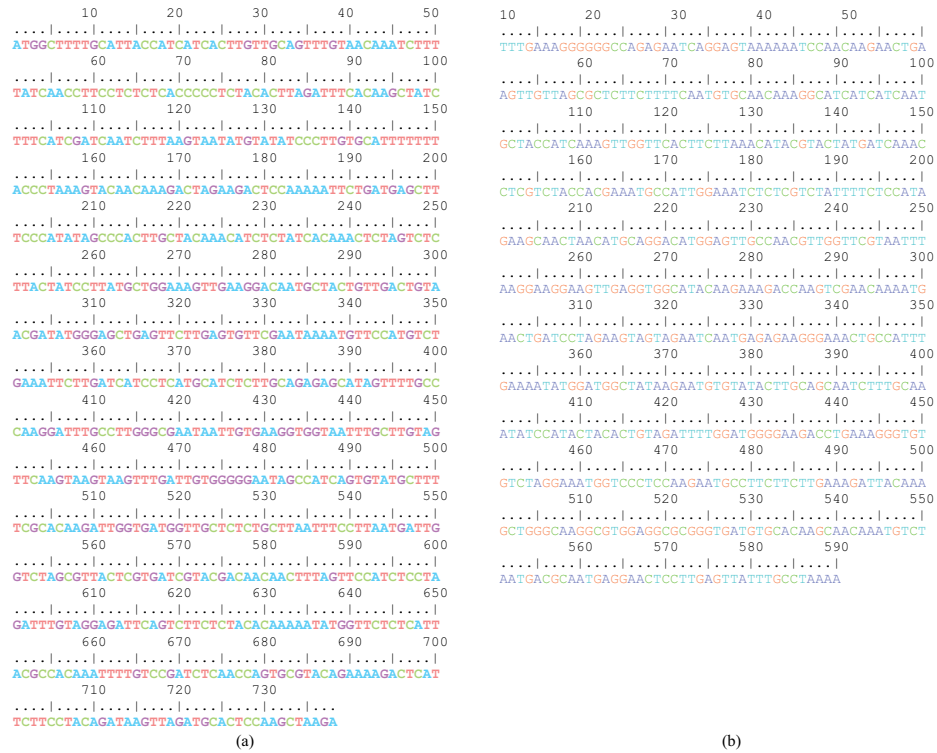


Figure 1: Sekuen *Pun1* gen from *C. frutescens* cv. Cakra Hijau isolated using primers: (a) F1/R1; (b) F2/R2.

3. Result and Discussion

In this study, combination primer forward and reverse F1/R1 primer obtained 738 bp exon-1 meanwhile combination primer forward and reverse F2/R2 primer obtained 590 bp exon-2 (Figure 1).

BLASTN analysis result that query coverage of F1/R1 Sequence is 100% from total sequence exon-1 *Pun1* gene from *C. frutescens* cv. Shuanla and *C. frutescens* cv. BG2814-6, and have 98% similarity sequence with exon-1 *Pun1* gene from *C. frutescens* cv. Shuanla and *C. frutescens* cv. BG2814-6 (Figure 2).

BLASTN analysis result that query coverage of F2/R2 Sequence is 99% from total sequence exon-2 *Pun1* gene from *C. frutescens* cv. Shuanla and *C. frutescens* cv. BG2814-6 and have 98% and 97% similarity sequence with exon-1 *Pun1* gene from *C. frutescens* cv. Shuanla and *C. frutescens* cv. BG2814-6, respectively (Figure 3).

F1/R1 sequence was analyzed with SIXFRAME (www.seqtool.edu) showing first open reading frame (ORF) is the most possible reading frame of the target gene because it contained start codon in the beginning of fragment (Figure 4). Exon-1 from *Pun1* gene containing a start codon [9].

Open reading frame analysis from F2/R2 sequence show that second ORF is the most possible reading frame of the target gene because it just have one stop codon

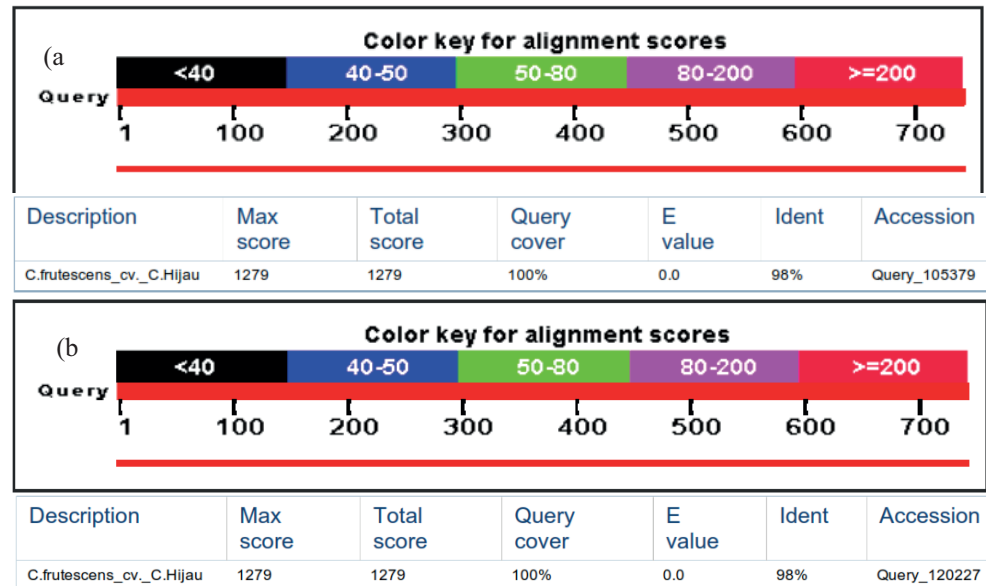


Figure 2: BLASTN analysis result of F1/R1 sequence *Pun1* gene from *C. frutescens* cv. Cakra Hijau compared with *Pun1* gene from: (a) *C. frutescens* cv. BG2814-6 (b) *C. frutescens* cv. Shuanla.

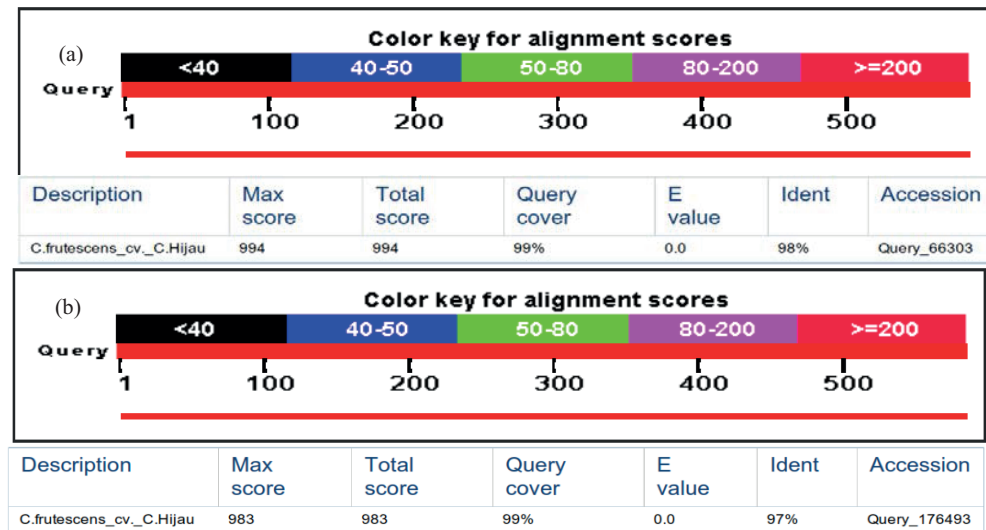


Figure 3: BLASTN analysis result of F2/R2 sequence *Pun1* gene from *C. frutescens* cv. Cakra Hijau compared with *Pun1* gene from: (a) *C. frutescens* cv.; BG2814-6; (b) *C. frutescens* cv. Shuanla.

in the end of sequence and not found another stop codon in the middle of sequence (Figure 5). Exon-2 *Pun1* gene containing a stop codon [11].

Sequences from exon-1 and exon-2 *Pun1* gene from *C. frutescens* cv. Cakra Hijau was analyzed using ClustalX to align amino acid sequence. The result of this analysis revealed that exon-1 *Pun1* gene of *C. frutescens* cv. Cakra Hijau located in the 1st up to 246th amino acid sequence of *C. frutescens* cv. BG2814-6 and *C. frutescens* cv. Shuanla (Figure 6).

```

M A F A L P S S L V A V C N K S F I X P
1 atggcTTTTgcattaccatcatcacttgTTgcagTTTgtaacaaatcttttatcaancct 60
S S L T P S T L R F H K L S F I D Q S L
61 tcctctcaccCCctctacacttagatTtcacaagctatctttcatcgatcaatcttta 120
S N M Y I P C A F F Y P K V Q Q R L E D
121 agtaatatgtatatcccttgTgcattTTTTtacctaagTacaacaagactagaagac 180
S K N S D E L S H I A H L L Q T S L S Q
181 tccaaaaattctgatgagctttcccatatagcccacttgctacaaacatctctatcacia 240
T L V S Y Y P Y A G K L K D N A T V D C
241 actctagtctcttactatccttatgctggaaagTgaaggacaatgctactgttgactgt 300
N D M G A E F L S V R I K C S M S E I L
301 aacgatatgggagctgagTtcttgagTtctgaataaaaatgTtccatgTctgaaatctt 360
D H P H A S L A E S I V L P K D L P W A
361 gatcatcctcatgcatctcttgcagagagcatagTtttgcccaaggattTgcctTggggc 420
N N C E G G N L L V V Q V S K F D C G G
421 aataattgtgaaggTggaattTgctTgtagTtcaagtaagTaatTgattgtggggga 480
I A I S V C F S H K I G D G C S L L N F
481 atagccatcagTgtatgcttttcgcacaagattggTgatggTtctctgcttaatttc 540
L N D W S S V T R D R T T T T L V P S P
541 cttaatgattggTctagcgttactcgtgatcgtacgacaacaactTtagTtccatctcct 600
R F V G D S V F S T Q K Y G S L I T P Q
601 agattTtaggagattcagTcttctctacacaaaaatagTtctctcattacgccacaa 660
I L S D L N Q C V Q K R L I L P T D K L
661 atTTgtccgatctcaaccagTgcgtacagaaaagactcattcttctacagataagTta 720
D A L Q A K
721 gatgactccaagctaaga 739

```

Figure 4: The first ORF F1/R1 sequence *Pun1* gene of *C. frutescens* cv. Cakra Hijau. M: Methionin or Start codon.

```

F E R G A R E S G V K N P T R T E V V S
2 ttTgaaagggggccagagaatcaggagTaaaaaatccaacaagaactgaagTtTtagc 61
A L L F X C A T K A S S S M L P S K L V
62 gctcttcttttcaantgtgcaacaaagTcatcatcaatgctaccatcaaagTtggtt 121
H F L N I R T M I K P R L P R N A I G N
122 cacttcttaacatacgtactatgatcaaaactcgtctaccacgaaatgccattggaaat 181
L S S I F S I E A T N M Q D M E L P T L
182 ctctcgtctatTTtctccatagaagcaactaacatgcaggacatggagTtgccaacgTtg 241
V R N L R K E V E V A Y K K D Q V E Q N
242 gTtTcgaatTtaaggaaggaagTtgagTggcatacaagaaagaccaagTcgaacaaaat 301
E L I L E V V E S M R E G K L P F E N M
302 gaactgatcctagaagtagtagaatcaatgagagaagggaaactgccattTgaaatag 361
D G Y K N V Y T C S N L C K Y P Y Y T V
362 gatggctataagaatgtgtatactTgcagcaactTTtgcaaatcactactactgta 421
D F G W G R P E R V C L G N G P S K N A
422 gattTtgatggggaagacctgaaagggTgtgtctaggaatggTccctccaagaatgcc 481
F F L K D Y K A G Q G V E A R V M X H K
482 ttcttctgaaagattacaaagctgggcaagggcgtggaggcggggtgatgTgcacaag 541
Q Q M S N X X R N E E L L E L F A *
542 caacaaatgtctaatgnnnnacgcaatgaggaactccttgagTtattTgcctaaaa 597

```

Figure 5: The second ORF F1/R1 sequence *Pun1* gene of *C. frutescens* cv. Cakra Hijau.*: Stop codon.

Analyzed of exon-2 *Pun1* gene from *C. frutescens* cv. Cakra Hijau show that sequence located in the 240th up to 440th amino acid sequence of *C. frutescens* cv. BG2814-6 and *C. frutescens* cv. Shuanla (Figure 7).

Alignment analysis showed that there was over lapping amino acid in *C. frutescens* cv. Cakra Hijau from 240th to 246th amino acid. Further study is needed to confirm this result.

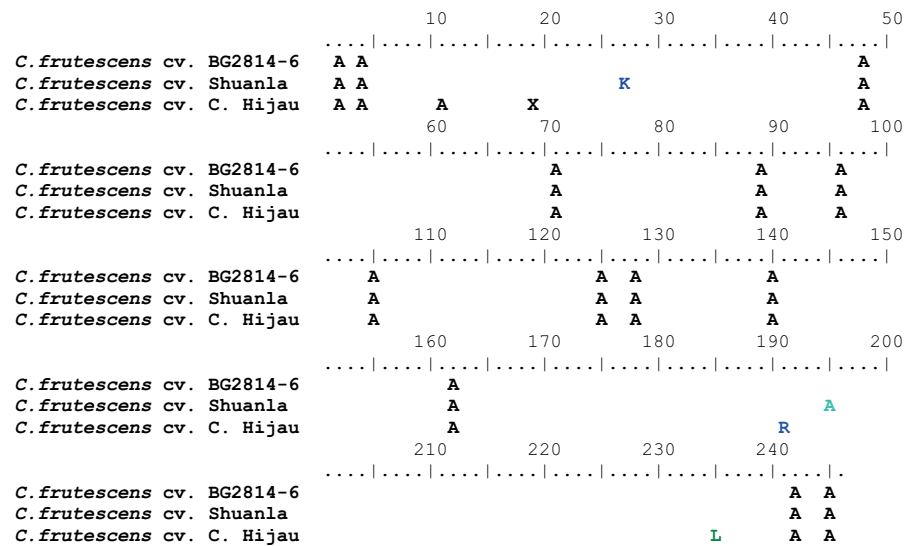


Figure 6: Aligenment of amino acid sequence from exon-1 *Pun1* gene *C. frutescens* cv. Cakra Hijau with *C. frutescens* cv. BG2814-6 and *C. frutescens* cv. Shuanla.

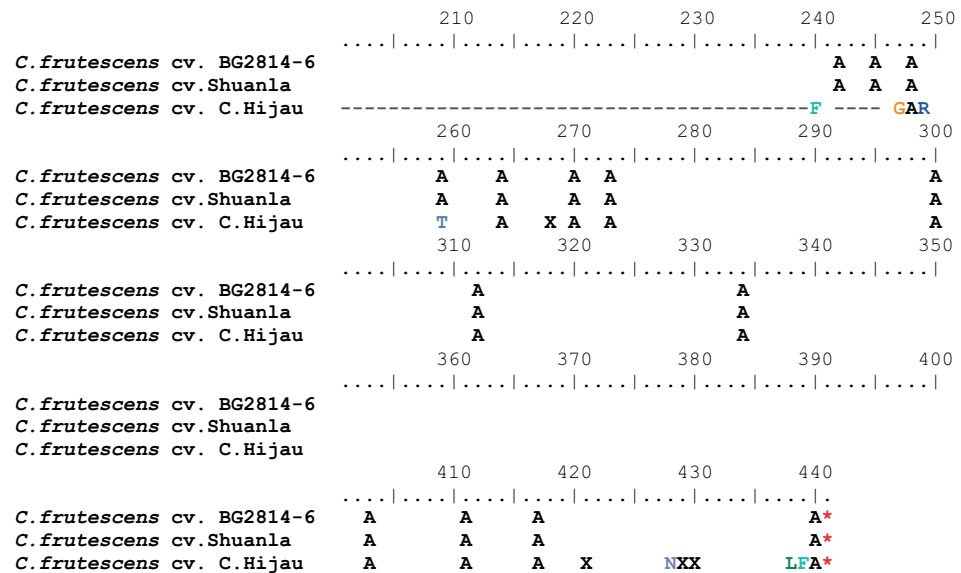


Figure 7: Alignment of amino acid from exon-2 *Pun1* gene *C. frutescens* cv. Cakra Hijau with *C. frutescens* cv. BG2814-6 and *C. frutescens* cv. Shuanla. .*: Stop codon.

4. Conclusions

This study has successfully isolated 738 bp length of exon-1 and 590 bp length of exon-2 of *Pun1* gene from *C. frutescens* cv. Cakra Hijau. Further study is needed because there was an over lapped amino acid by 6 bp length.

Acknowledgements

The author would like to acknowledge to Dahlia, Heni Edrawati, Ayu Linda Febriani, Riska Anggraini, and Muthia Naila Mazieda for technical assistance and useful discussion. This work was supported in part by PKM-PE DIKTI 2015 and Biotechnology Laboratory State University of Malang.

References

- [1] C. B. Heiser and P. G. Smith, "The cultivated capsicum peppers," *Economic Botany*, vol. 7, no. 3, pp. 214-227, 1953.
- [2] T. Djarwaningsih, *Capsicum spp. (cabai): asal, persebaran dan nilai ekonomi. [Capsicum spp. (chili): origin, distribution and economic value]* *Biodiversitas*, vol. 6, persebaran dan nilai ekonomi. [Capsicum spp. (chili, origin, 2005, in Bahasa Indonesia.
- [3] Rukmana., "Usaha tani cabai rawit. [Farm of chili]," in *Rukmana. Usaha tani cabai rawit. [Farm of chili]*, p. 13, Kanikus, Yogyakarta, 2002, 20 in Bahasa Indonesia.
- [4] E. Zamski, O. Shoham, D. Palevitch, and A. Levy, "Ultrastructure of Capsaicinoid-Secreting Cells in Pungent and Nonpungent Red Pepper (*Capsicum annuum* L.) Cultivars," *Botanical Gazette*, vol. 148, no. 1, pp. 1-6, 1987.
- [5] H. Fujiwake, T. Suzuki, and K. Iwai, "Capsaicinoid formation in the protoplast from the placenta of capsicum fruits," *Agricultural and Biological Chemistry*, vol. 46, no. 10, pp. 2519-2592, 1982.
- [6] K. Iwai, T. Suzuki, and H. Fujiwake, "Formation and accumulation of pungent principle of hot pepper fruits, capsaicin and its analogues, in capsicum annuum var. Annuun cv. karayatsubusa at different growth stages after flowering," *Agricultural and Biological Chemistry*, vol. 43, no. 12, pp. 2493-2498, 1979.
- [7] J. Curry, M. Aluru, M. Mendoza, J. Nevarez, M. Melendrez, and M. A. O'Connell, "Transcripts for possible capsaicinoid biosynthetic genes are differentially accumulated in pungent and non-pungent *Capsicum* spp," *Plant Science*, vol. 148, no. 1, pp. 47-57, 1999.
- [8] M. Kim, S. Kim, S. Kim, and B.-D. Kim, "Isolation of cDNA Clones Differentially Accumulated in the Placenta of Pungent Pepper by Suppression Subtractive Hybridization," *Molecules and Cells*, vol. 11, no. 2, pp. 213-219, 2001.
- [9] C. Stewart Jr., B.-C. Kang, K. Liu et al., "The Pun1 gene for pungency in pepper encodes a putative acyltransferase," *Plant Journal*, vol. 42, no. 5, pp. 675-688, 2005.
- [10] C. Stewart Jr., M. Mazourek, G. M. Stellari, M. O'Connell, and M. Jahn, "Genetic control of pungency in *C. chinense* via the Pun1 locus," *Journal of Experimental Botany*, vol. 58, no. 5, pp. 979-991, 2007.
- [11] G. M. Stellari, M. Mazourek, and M. M. Jahn, "Contrasting modes for loss of pungency between cultivated and wild species of *Capsicum*," *Heredity*, vol. 104, no. 5, pp. 460-471, 2010.

- [12] M. Habibi, A. Madhahah Manggabarani, E. Sri Sulasmi, and D. Listyorini, "AT3 (Acyltransferase) Gene Isolated From *Capsicum frutescens* cv. Cakra Hijau," *Journal of Tropical Life Science*, vol. 3, no. 2, pp. 83–86, 2013.
- [13] AM. Manggabarani, *Isolasi fragmen bagian depan dan tengah kedua gen acyltransferase (AT3) dari Capsicum frutescens L. cv. CakraHijau. [Isolation fragment front and center of both genes acyltransferase (AT3) from Capsicum frutescens L. cv. CakraHijau] [Undergraduate thesis], Malang: Universitas Negeri Malang, ., [in Bahasa Indonesia], Universitas Negeri Malang, Malang, 2013.*